RESEARCH

Open Access



Effects of L-carnitine supplementation on lipid profiles in patients with coronary artery disease

Bor-Jen Lee¹, Jun-Shuo Lin², Yi-Chin Lin³ and Ping-Ting Lin^{3,4*}

Abstract

Background: L-carnitine (LC) plays an important physiologic role in lipid metabolism. To date, no clinical study has been performed to examine the effect of LC supplementation on the lipid status of coronary artery disease (CAD) patients. The aim of this study was to investigate the lipid lowering effects of LC supplementation (1000 mg/d) in CAD patients.

Methods: CAD patients were identified by cardiac catheterization as having at least 50 % stenosis of one major coronary artery. Forty-seven subjects were recruited and randomly assigned to the placebo (n = 24) and to the LC (n = 23) groups. The intervention was administered for 12 weeks. The levels of LC, lipid profiles, and antioxidant enzyme activity (superoxide dismutase, SOD) were measured.

Results: The subjects in the LC group had significantly higher SOD activity (20.7 ± 4.2 versus 13.1 ± 2.9 U/mg of protein, P < 0.01), high density lipoprotein-cholesterol (1.34 ± 0.42 vs. 1.16 ± 0.24 mmol/L, HDL-C, P = 0.03), and apolipoprotein-A1 (Apo-A1, 1.24 ± 0.18 vs. 1.12 ± 0.13 g/L, P = 0.02) than those in the placebo group at week 12. Triglyceride (TG) level was slightly significantly reduced (1.40 ± 0.74 vs. 1.35 ± 0.62 mmol/L, P = 0.06) and the level of LC was negatively correlated with TG and apolipoprotein-B (Apo-B), and positively correlated with HDL-C and Apo-A1 after LC supplementation. Additionally, SOD activity was significantly negatively correlated with lipid profiles (total cholesterol, TG, and Apo-B) after supplementation.

Conclusion: LC supplementation at a dose of 1000 mg/d showed significantly increased in HDL-C and Apo-A1 levels and a slight decrease in TG levels but no other changes in other lipids in CAD patients, and this lipid-lowering effect may be related to its antioxidant ability. Further studies should be conducted to define an optimal dose of LC for lipid-lowering in patients with CAD.

Trial registration: Clinical Trials.gov Identifier: NCT01819701

Keywords: L-carnitine supplement, Lipid profiles, Antioxidant, Coronary artery disease

Background

L-carnitine (LC) plays an important physiologic role in lipid metabolism. LC may carry long-chain fatty acids across the inner mitochondrial membrane for β -oxidation and adenosine triphosphate (ATP) production [1, 2]. Recent studies have demonstrated that LC is not only a transporter of lipids but is also an

antioxidant [3, 4]. LC appears to interfere with the formation of reactive oxygen species, through its ferrous ion metal-chelating ability, thus stabilizing the free radicals formed on the α -carbon by conjugation [4, 5]. In addition to its antioxidant activity, LC also exhibits anti-inflammatory properties [6–8]. LC may inhibit the nuclear factor-kappa B pathway through the suppression of reactive oxygen species formation [9–11]. Whereas coronary artery disease (CAD) is associated with higher levels of oxidative stress and inflammation and hyperlipidemic status [12], it may be helpful to administer LC to CAD patients given its physiological functions.



© 2016 The Author(s). **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

^{*} Correspondence: apt810@csmu.edu.tw

³School of Nutrition, Chung Shan Medical University, Taichung 40201, Taiwan ⁴Department of Nutrition, Chung Shan Medical University Hospital, Taichung 40201, Taiwan

Full list of author information is available at the end of the article

A number of clinical studies have been conducted to examine the lipid-lowering effects of LC supplementation in patients with renal disease [7, 8, 12–17], diabetes [18–20], or hyperlipidemia [21, 22], however, the results regarding the lipid-lowering effects of LC supplementation are inconsistent due to different disease statuses. To date, no clinical study has been performed to examine the effect of LC supplementation on the lipid status of CAD patients. Therefore, the purpose of this study was to investigate the effect of LC supplementation (1000 mg/d) on lipid levels in CAD patients.

Methods

Study design

A single-blind, randomized, parallel, placebo-controlled trial was conducted. CAD patients were diagnosed by cardiac catheterization (at least 50 % stenosis of one major coronary artery) or by percutaneous transluminal coronary angioplasty (PTCA) for stable condition. We excluded the patients with diabetes, liver, or renal diseases, along with those undergoing acenocoumarol, thyroid hormone, warfarin, and a high dose of statin therapy [Atorvastatin (Lipitor) or Rosuvastatin (Crestor), 30-40 mg/d] or currently using vitamin supplements. The study was approved by the Institutional Review Board of Taichung Veterans General Hospital, Taiwan and the clinical trial was registered at Clinical Trials.gov (NCT01819701). This clinical trial started recruiting subjects at January 2013, and the last subject was completed in February 2014. Each subject provided written informed consent to participate in the study. A total of 47 CAD patients were recruited to this study and randomly assigned to the placebo (n = 24) or to the LC (1000 mg/day, n = 23) group. The supplements, placebo (starch) and LC capsules were provided by New Health Taiwan Co., Ltd (Taichung, Taiwan). At the beginning of the study, the investigators instructed the subjects to take two capsules daily (LC supplements 500 mg b.i.d). To monitor the subject compliance, we asked the subjects to return the supplied bag of capsules every 4 weeks to verify the capsule count. The intervention was administered for 12 weeks. The sampling and trial profiles along with the number of subjects who completed the study in each group are presented in Fig. 1.

At baseline, we measured the subjects' blood pressure, body weight, height, and waist circumference, and then calculated the body mass index (kg/m²). General obesity was defined as body mass index \geq 27 kg/m² [23], and central obesity was defined as waist circumferences \geq 90 cm. Dietary intakes were assessed using 24-h recall dietary records at baseline. The dietary records were analyzed using the Nutritionist Professional software package (E-Kitchen Business Corp., Taiwan).



Lipid profiles, L-carnitine, and antioxidant enzyme activity measurements

The fasting blood specimens were collected in vacutainer tubes without anticoagulant (Becton Dickinson, Rutherford, NJ, USA). The samples were centrifuged at 3,000 rpm for 15 minutes at 4 °C, and the serum was separated. Serum total cholesterol (TC), triacylglycerol (TG), low density lipoprotein-cholesterol (LDL-C), and high density lipoprotein-cholesterol (HDL-C) were measured using an automated biochemical analyzer (Hitachi-7180E, Tokyo, Japan). The levels of apolipoprotein A-1 (Apo-A1) and apolipoprotein-B (Apo-B) were measured using the polyethylene glycerol (PEG) enhanced immunoturbidimetric assay method (Siemens Healthcare Diagnostics Inc., New York, USA). The level of LC was measured by the enzyme-linked immunosorbent assay (ELISA) method using commercially available kits (Cusabio, Wuhan, China) according to the instructions made available by the suppliers.

The antioxidant enzyme activity (superoxide dismutase, SOD) were measured in the fresh red blood cells (RBCs) samples. The RBCs were washed with normal saline after removing the plasma, and then diluted with 25x sodium phosphate buffer for measurement. The methods for measuring SOD activity have been described previously [24]. The protein content of the plasma and RBCs was determined based on the biuret reaction of the bicinchoninic acid assay (BCA kit, Thermo, Rockford, IL, USA). SOD values were expressed as U/mg of protein. All of the analyses were performed in duplicate.

Statistical analysis

Means and standard deviations (s.d.) were calculated for all data. Kolmogorov-Smirnov test was used to examine the normal distribution of variables. Student's t-test or the Mann-Whitney rank sum test was used to compare mean values for continuous variables between the placebo and LC groups. Paired t-test or Wilcoxon signed rank test was used to compare the data before and after supplementation within the group. For categorical response variables, differences between the two groups were assessed by the Chi-square test or Fisher's exact test. Simple linear regressions were used to examine the correlations between the levels of LC (as a dependent variable) and lipid profiles after supplementation. Pearson correlations were used to examine the correlations between the antioxidant enzyme activity and lipid profiles after LC supplementation and the correlations between the changes (week 12-0, ${\scriptscriptstyle \Delta})$ in antioxidant enzyme activity and lipid profiles. Statistical significance was set at P < 0.05. All statistical analyses were performed using SigmaPlot software (version 12.0, Systat, San Jose, CA, USA).

Results

Characteristics of the subjects at baseline

A total of 39 CAD subjects completed the 12-week interventional study (Placebo, n = 19; LC, n = 20). The baseline characteristics of the subjects are shown in Table 1. The means and s.d. for age, blood pressure, and dietary intakes as well as the percentages of general obesity and central obesity were not significantly different between the two groups at baseline.

Effects of LC supplementation on the levels of LC, SOD activity, and lipid profiles

The subjects in the LC group had significantly increased LC level (40.0 ± 12.0 versus.35.2 ± 12.0 µmol/L, P = 0.02) and SOD activity (20.7 ± 4.2 versus 13.1 ± 2.9 U/mg of protein, P < 0.01) compared with those in the placebo group at week 12 as well as after LC supplementation (LC, 33.6 ± 13.6 to 40.0 ± 12.0 µmol/L, P = 0.04; SOD, 14.8 ± 2.9 to 20.7 ± 4.2 U/mg of protein, P < 0.01). The lipid profiles after supplementation are shown in Fig. 2. The subjects in the LC group had significantly higher level of HDL-C (1.34 ± 0.42 vs. 1.16 ± 0.24 mmol/L, P = 0.03) and Apo-A1 (1.24 ± 0.18 vs. 1.12 ± 0.13 g/L, P = 0.02) than those in the placebo group at week 12. After

Table 1 Baseline characteristics of the subj	ects
--	------

	Placebo (<i>n</i> = 19)		LC (n = 20)	
	Mean	s.d.	Mean	s.d
Gender (male)	19		20	
Age (y)	72.7	10.1	71.9	10.6
SBP (mmHg)	127.3	6.0	128.4	10.4
DBP (mmHg)	74.5	4.4	72.3	4.7
General obesity ^a (%)	36.8		30.0	
Central obesity ^a (%)	84.2		65	
Smoking ^b (n, %)	3 (15.8 %)		4 (20.0 %)	
Drinking ^c (n, %)	3 (15.8 %)		3 (15.0 %)	
Dietary intake				
Energy (kcal/day)	1501.4	63.7	1534.2	41.6
Protein (g/day, % of total calories)	54.2 (14.2)	11.2	51.1 (12.9)	6.9
Fat (g/day, % of total calories)	43.3 (26.4)	9.7	43.2 (23.8)	10.6
SFA (g/day)	11.7	5.3	10.6	5.4
PUFA (g/day)	14.0	3.5	15.9	3.6
MUFA (g/day)	13.3	3.8	13.0	4.9
Carbohydrate (g/day, % of total calories)	225.3 (60.3)	16.3	236.7 (63.5)	24.7

^aGeneral obesity was defined as body mass index $\ge 27 \text{ kg/m}^2$ and central obesity was defined as waist circumferences $\ge 90 \text{ cm}$. ^bSmoking: the individual currently smokes one or more cigarettes per day. ^cDrinking: the individual regularly drinks one or more drinks per day

DBP diastolic blood pressure; *LC* L-carnitine; *MUFA* monounsaturated fatty acids; *PUFA* polyunsaturated fatty acids; *SBP* systolic blood pressure; *SFA* saturated fatty acids

Lee et al. Lipids in Health and Disease (2016) 15:107



LC supplementation, the level of TG was slightly reduced but did not achieve statistical significance $(1.40 \pm 0.74 \text{ vs.} 1.35 \pm 0.62 \text{ mmol/L}, P = 0.06$, respectively). However, no significant change in the levels of TC, LDL-C, and Apo-B after 12 weeks of LC supplementation. Notably, the level of Apo-A1 was decreased in the placebo group after 12 weeks

intervention $(1.20 \pm 0.12 \text{ vs.} 1.12 \pm 0.13 \text{ g/L}, P = 0.05)$; however, the result was not statistically significant.

Correlations between the levels of lipid profiles and LC after supplementation

The correlations between the levels of lipid profiles and LC after supplementation and these results are

shown in Fig. 3. The level of LC was significantly negatively correlated with TG (β = -0.14, *P* = 0.01). Although not statistically significant, LC level correlated with Apo-B (β = -0.02, *P* = 0.08), HDL-C (β = 0.07, *P* = 0.07) and Apo-A1 (β = 0.01, *P* = 0.09) after 12 weeks of supplementation.

Correlations between the SOD activity and lipid profiles after LC supplementation

The correlations between the antioxidant enzyme activity and lipid profiles after LC supplementation are shown in Table 2. SOD activity was significantly negatively correlated with TC (r = -0.34, P = 0.03), TG (r = -0.53, P < 0.01), and Apo-B (r = -0.32, P < 0.05) after 12 weeks of LC supplementation. Furthermore, we assessed the correlations between the changes (week 12-0, \triangle) in antioxidant enzyme activity and lipid profiles (Table 3). There was a significant correlation between the changes level of SOD activity and the changes in TC (r = -0.41, P < 0.01), LDL-C (r = -0.45, P < 0.01), and Apo-B (r = -0.32, P < 0.01).

Discussion

Significant lipid-lowering effects of LC supplementation have been demonstrated in hemodialysis patients [16, 17]. Naini et al. treated chronic hemodialysis patients with oral LC supplementation at doses of 750 mg/d and 1000 mg/d for 8 weeks and 16 weeks, respectively. LC supplementation at a dose of 750 mg/d significantly decreased levels of lipid (TC, TG, and LDL-C) [17], whereas LC supplementation at a dose of 1000 mg/d significantly decreased levels of TC and TG and increased the level of HDL-C [16]. LC may increase the mitochondrial transport of fatty acids and reduce fatty acid availability for lipid synthesis [25]. LC has also improved the levels of lipid in CAD given that LC is a lipid transporter. In this study, we observed that LC administered at a dose of 1000 mg/d for 12 weeks correlated with decreased lipid profile (TG) levels and increased levels of HDL-C and Apo-A1. Our previous studies demonstrated that LC supplementation at a dose of 1000 mg/d was significantly associated with increased



Table 2 Correlations between the antioxidant enzyme activity and lipid profiles after L-carnitine supplementation

	SOD (U/mg protein)
	r ^a (P values)
TC (mmol/L)	-0.34 (0.03)
TG (mmol/L)	-0.53 (< 0.01)
LDL-C (mmol/L)	-0.15 (0.36)
HDL-C (mmol/L)	0.11 (0.52)
Apo-A1 (g/L)	-0.18 (0.29)
Apo-B (g/L)	-0.32 (< 0.05)

^acorrelation coefficients

Apo apolipoprotein; HDL-C high density lipoprotein-cholesterol; LC L-carnitine; LDL-C low density lipoprotein-cholesterol; SOD superoxide dismutase; TC total cholesterol; TG triglycerides

antioxidant capacity in CAD patients [26]. In addition, the level of LC was significantly positively correlated with antioxidant enzymes activity (SOD, $\beta = 0.72$, P < 0.01) and reduced oxidative stress and inflammatory status in the patients with CAD after supplementation [26, 27]. In this study, we observed no significant correlation between LC and lipids levels or between SOD activity and lipid levels at baseline. Instead, antioxidant enzyme activity (SOD) was significantly negatively correlated with lipid levels (TC, TG, and Apo-B) after LC supplementation (Table 2). As a result, wehypothesize that the effect of LC supplementation on lipids (TG, HDL-C, and Apo-A1) may be related to its antioxidant capacity.

Apolipoproteins, such as Apo-B and Apo A-1, are the major apolipoproteins associated with LDL-C and HDL-C, respectively [28]. In this study, we observed a significant increase in the levels of Apo-A1 and HDL-C, and the level of LC was slightly negatively correlated with Apo-B after supplementation. However, no significant change in the levels of TC and LDL-C were noted after LC supplementation. In a clinical study, Malaguarnera et al. investigated diabetes patients using a higher dose of

Table 3 Correlations between the changes (week 12-0, \triangle) in antioxidant enzyme activity and lipid profiles

	△SOD (U/mg protein)	
	r ^a (P values)	
△TC (mmol/L)	-0.41 (< 0.01)	
△TG (mmol/L)	-0.11 (0.34)	
△LDL-C (mmol/L)	-0.45 (< 0.01)	
△HDL-C (mmol/L)	-0.04 (0.77)	
△Apo-A1 (g/L)	-0.09 (0.44)	
△Apo-B (g/L)	-0.32 (< 0.01)	

^acorrelation coefficients

Apo apolipoprotein; HDL-C high density lipoprotein-cholesterol; LC L-carnitine; LDL-C low density lipoprotein-cholesterol; SOD superoxide dismutase; TC total cholesterol; TG triglycerides LC supplementation of 2000 mg/d for 12 weeks and observed significant decreases in the levels of TC, TG, LDL-C, oxidized LDL-C, and Apo-B, and increases in the levels of HDL-C and Apo-A1 [20]. Hyperlipidemia is a contributor to CAD; thus, we considered that if the disease has already developed, the lipid-lowering effect of LC supplementation at a dose of 1000 mg/d may not be sufficient. Although LC at a dose of 1000 mg/d can significantly increase the antioxidant activity [26], a higher dose of LC (>1000 mg/d) administration could be considered in anti-hyperlipidemic strategies [29, 30].

In the present study, TG level decreased after supplementation, but the reduction did not achieve statistical significance, which may have been attributed to the stable CAD subjects and the limited number of subjects (Placebo, n = 13; LC, n = 8) with hyperlipidemia (a ratio of TC to HDL \geq 5.0, TG \geq 1.7 mmol/L, or LDL-C \geq 3.4 mmol/L) [31]. Due to the small number of subjects having hyperlipidemia in the LC group, the detection of the lipid-lowering effects in these subjects may have been limited. Because higher oxidative stress and inflammation status are early events in the evolution of hyperlipidemia [32, 33], we suggest that an antioxidant supply (such as LC) may help prevent or delay the development of CAD [26, 27]. The lipid-lowering effects of a clinically relevant degree of LC supplementation might be related to its dosage. Further studies should be conducted to define an optimal dose of LC for lipidlowering in patients with CAD and explore whether it could be used as a nutraceutical adjuvant in the cases of dyslipidemia [34].

The strength of this study is that it is the first clinical study to investigate the lipid-lowering effects of LC supplementation in CAD patients, and we measured the levels of LC, lipid profiles, and antioxidant enzyme activity. This study provides direct evidence to clarify the relationship among the levels of LC, lipid, and antioxidant ability; however, longer intervention studies with larger sample sizes should be performed to confirm the lipid-lowering effects after LC intervention in patients with hyperlipidemia. There are some limitations of the present study that should be mentioned. First, the number of participants was small; however, we performed the post hos calculations to examine the statistical power for lipid profiles. The statistical power of the differences of TG, HDL-C, and Apo-A1 were 0.97, 0.86, and 0.86, respectively. Second, a few of the subjects had hyperlipidemia in the LC group; thus, the detection of the lipid-lowering effects in these subjects may have been limited. Moreover, an optimal dose of LC for lipid-lowering should be defined. Further studies should also investigate the effect of LC on NADPH oxidase activity, HMG-CoA reductase activity and TG synthesis to clarify if LC lowers lipids by enhancing

mitochondrial fatty acid oxidation or by reducing oxidative stress.

Conclusion

In conclusion, LC supplementation at a dose of 1000 mg/d exhibited significant increases in HDL-C and Apo-A1 levels and a slight decrease in TG levels, but no other changes in other lipids were noted in CAD patients. This lipid-lowering effect may be related to its antioxidant ability.

Authors' contributions

BJL performed the study, performed the data analyses, and drafted the manuscript. JSL and YCL performed the study and sample analyses. PTL conceived the study, participated in its design, and coordination, and helped to draft the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Acknowledgements

This study was supported by a grant from the Chung Shan Medical University (CSMU-INT-101-09), Taiwan. We express our sincere appreciation to the subjects for their participation and to Dr. Hsia, who kindly provided the supplements for this trial. We thank the nurses at Taichung Veterans General Hospital for providing expert assistance in blood sample collection and data analysis.

Author details

¹The Intensive Care Unit, Taichung Veterans General Hospital, Taichung 40705, Taiwan. ²Department of Nutrition, Sinying Hospital, Ministry of Health and Welfare, Tainan 73042, Taiwan. ³School of Nutrition, Chung Shan Medical University, Taichung 40201, Taiwan. ⁴Department of Nutrition, Chung Shan Medical University Hospital, Taichung 40201, Taiwan.

Received: 18 February 2016 Accepted: 15 June 2016 Published online: 17 June 2016

References

- Kendler BS. Carnitine: an overview of its role in preventive medicine. Prev Med. 1986;15:373–90.
- Flanagan JL, Simmons PA, Vehige J, Willcox MD, Garrett Q. Role of carnitine in disease. Nutr Metab. 2010;7:30.
- Gülçin I. Antioxidant and antiradical activities of L-carnitine. Life sci. 2006;78: 803–11.
- Ribas GS, Vargas CR, Wajner M. L-carnitine supplementation as a potential antioxidant therapy for inherited neurometabolic disorders. Gene. 2014;533: 469–76.
- Kolodziejczyk J, Saluk-Juszczak J, Wachowicz B. L-Carnitine protects plasma components against oxidative alterations. Nutrition. 2011;27:693–9.
- Savica V, Santoro D, Mazzaglia G, Ciolino F, Monardo P, Calvani M, et al. L-carnitine infusions may suppress serum C-reactive protein and improve nutritional status in maintenance hemodialysis patients. J Ren Nutr. 2005; 15:225–30.
- Duranay M, Akay H, Yilmaz FM, Senes M, Tekeli N, Yücel D. Effects of L-carnitine infusions on inflammatory and nutritional markers in haemodialysis patients. Nephrol Dial Transplant. 2006;21:3211–4.
- Shakeri A, Tabibi H, Hedayati M. Effects of L-carnitine supplement on serum inflammatory cytokines, C-reactive protein, lipoprotein (a), and oxidative stress in hemodialysis patients with Lp (a) hyperlipoproteinemia. Hemodial Int. 2010;14:498–504.
- Cetinkaya A, Bulbuloglu E, Kantarceken B, Ciralik H, Kurutas EB, Buyukbese MA, et al. Effects of L-carnitine on oxidant/antioxidant status in acetic acidinduced colitis. Dig Dis Sci. 2006;51:488–94.
- Koc A, Ozkan T, Karabay AZ, Sunguroglu A, Aktan F. Effect of L-carnitine on the synthesis of nitric oxide in RAW 264A7 murine macrophage cell line. Cell Biochem Funct. 2011;29:679–85.
- Moeinian M, Ghasemi-Niri SF, Mozaffari S, Abdollahi M. Synergistic effect of probiotics, butyrate and L-Carnitine in treatment of IBD. JMHI. 2013;7:50–3.

- Stocker R, Keaney JF. Role of oxidative modifications in atherosclerosis. Physiol Rev. 2004;84:1381–478.
- Shojaei M, Djalali M, Khatami M, Siassi F, Eshraghian M. Effects of carnitine and coenzyme Q10 on lipid profile and serum levels of lipoprotein(a) in maintenance hemodialysis patients on statin therapy. Iran J Kidney Dis. 2011;5:114–8.
- Suchitra MM, Ashalatha VL, Sailaja E, Rao AM, Reddy VS, Bitla AR, et al. The effect of L-carnitine supplementation on lipid parameters, inflammatory and nutritional markers in maintenance hemodialysis patients. Saudi J Kidney Dis Transpl. 2011;22:1155–9.
- Tabibi H, Hakeshzadeh F, Hedayati M, Malakoutian T. Effects of I-carnitine supplement on serum amyloid A and vascular inflammation markers in hemodialysis patients: a randomized controlled trial. J Ren Nutr. 2011;21: 485–91.
- Naini AE, Moradi M, Mortazavi M, Amini Harandi A, Hadizadeh M, Shirani F, et al. Effects of oral L-carnitine supplementation on lipid Profile, anemia, and quality of life in chronic renal disease patients under hemodialysis: a randomized, double-blinded, placebo-controlled trial. J Nutr Metab. 2012; 2012;510483.
- Naini AE, Sadeghi M, Mortazavi M, Moghadasi M, Harandi AA. Oral carnitine supplementation for dyslipidemia in chronic hemodialysis patients. Saudi J Kidney Dis Transpl. 2012;23:484–8.
- Derosa G, Cicero AF, Gaddi A, Mugellini A, Ciccarelli L, Fogari R. The effect of L-carnitine on plasma lipoprotein(a) levels in hypercholesterolemic patients with type 2 diabetes mellitus. Clin Ther. 2003;25:1429–39.
- Rahbar AR, Shakerhosseini R, Saadat N, Taleban F, Pordal A, Gollestan B. Effect of L-carnitine on plasma glycemic and lipidemic profile in patients with type II diabetes mellitus. Eur J Clin Nutr. 2005;59:592–6.
- Malaguarnera M, Vacante M, Avitabile T, Malaguarnera M, Cammalleri L, Motta M. L-Carnitine supplementation reduces oxidized LDL cholesterol in patients with diabetes. Am J Clin Nutr. 2009;89:71–6.
- Hlais S, Reslan DR, Sarieddine HK, Nasreddine L, Taan G, Azar S. Effect of lysine, vitamin B(6), and carnitine supplementation on the lipid profile of male patients with hypertriglyceridemia: a 12-week, open-label, randomized, placebo-controlled trial. Clin Ther. 2012;34:1674–82.
- 22. Lombardo F, Lunghi R, Pallotti F, Palumbo A, Senofonte G, Cefaloni AC, et al. Effects of a dietary supplement on cholesterol in subjects with moderate hypercholesterolemia. Clin Ter. 2013;164:e147–50.
- Chen HJ, Bai CH, Yeh WT, Chiu HC, Pan WH. Influence of metabolic syndrome and general obesity on the risk of ischemic stroke. Stroke. 2006;37:1060–4.
- 24. Marklund S, Marklund G. Involvement of superoxide anion radical in autoxidation of pyrogallol and a convenient assay for superoxide dismutase. Eur J Biochem. 1974;47:469–74.
- Bartel LL, Hussey JL, Shrago E. Perturbation of serum carnitine levels in human adults by chronic renal disease and dialysis therapy. Am J Clin Nutr. 1981;34:1314–20.
- Lee BJ, Lin JS, Lin YC, Lin PT. Effects of L-carnitine supplementation on oxidative stress and antioxidant enzymes activities in patients with coronary artery disease: a randomized, placebo-controlled trial. Nutr J. 2014;13:79.
- Lee BJ, Lin JS, Lin YC, Lin PT. Anti-inflammation effects of L-carnitine supplementation (1000 mg/d) in coronary artery disease patients. Nutrition. 2015;31:475–9.
- Walldius G, Jungner I. Apolipoprotein B and apolipoprotein A-I: risk indicators of coronary heart disease and targets for lipid-modifying therapy. J Intern Med. 2004;255:188–205.
- 29. Brescia F, Balestra E, Iasella MG, Damato AB. Effects of combined treatment with simvastatin and L-carnitine on triglyceride levels in diabetic patients with hyperlipidaemia. Clin Drug Investig. 2002;22:23–8.
- Chen Y, Abbate M, Tang L, Cai G, Gong Z, Wei R, Zhou J, Chen X. L-Carnitine supplementation for adults with end-stage kidney disease requiring maintenance hemodialysis: a systematic review and meta-analysis. Am J Clin Nutr. 2014;99:408–22.
- National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). Third Report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III) final report. Circulation. 2002;106:3143–421.
- 32. Yang RL, Shi YH, Hao G, Li W, Le GW. Increasing oxidative stress with progressive hyperlipidemia in human: relation between malondialdehyde and atherogenic Index. J Clin Biochem Nutr. 2008;43:154–8.

- Adekunle AS, Adedeji AL, Oyewo EO, Adedosu OT, Omotoso AT. Hyperlipidemia induced by atherogenic diet enhanced oxidative stress in the kidney and inflammatory responses: an in-vivo study. Asian J Natural Applied Sci. 2013;2:82–93.
- Scicchitano P, Cameli M, Maiello M, Modesti PA, Muiesan ML, Nova S, et al. Nutraceuticals and dyslipidaemia: Beyond the common therapeutics. J Funct Foods. 2014;6:11–32.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at www.biomedcentral.com/submit



BioMed Central